

## Metal Complexes of the Antibiotic Nalidixic Acid

NORAH BARBA BEHRENS\*, GUILLERMO MENDOZA DIAZ

*Departamento de Química Inorganica, Facultad de Química, U.N.A.M., 04510 Mexico D.F., Mexico*

and DAVID M. L. GOODGAME

*Chemistry Department, Imperial College of Science and Technology, London SW7 2AY, U.K.*

(Received February 18, 1986)

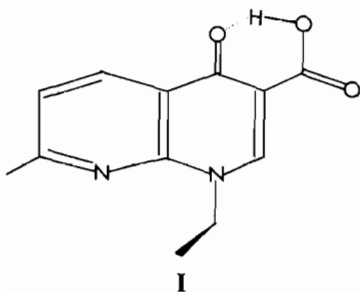
### Abstract

The preparations and spectral properties are reported of a range of complexes of nalidixic acid (= HNaI) with some metal ions in the series Cr–Zn and also Mg, Ca, Cd, Hg and Pd(II). Most of the compounds formed by the divalent metal ions had a 2:1 HNaI:metal ratio, and some of them are polymeric. Complexes in which the carboxylate group of HNaI functions as a chelate were isolated with Cu(II), Pd(II), Fe(III) and Cr(III).

### Introduction

Metal ions play a vital role in a vast number of widely differing biological processes. The interaction of these ions with drugs administered for therapeutic reasons is a subject of considerable interest. It is known that some drugs act via chelation [1] or by inhibiting metalloenzymes [2] but for most of the drugs that could act as potential ligands little is known about how metal binding influences their activity.

In order to enhance understanding of drug–metal ion interactions, we have been studying the complexing ability of nalidixic acid (HNaI) (I) and its analogues. These antibiotics are used in the clinical



treatment of urinary tract infections caused by Gram-negative bacteria [3].

The antibacterial activity of nalidixic acid is apparently due to inhibition of DNA synthesis. Nalidixic acid is an inhibitor of the DNA-gyrase, a topoisomerase having a tetrameric structure formed by two subunits, A and B, and it is believed that subunit A is the direct target of the drug [4, 5]. Unfortunately the composition of the target is not yet known in detail, but it has been suggested that it may be a metalloenzyme [6].

A recent study [7] of the mechanism of action of quinolone antibacterial agents (nalidixic acid being the first member of this family) suggested that they do not bind to gyrase subunit A, but rather to DNA. In this hypothesis, the proximal inhibitor of DNA gyrase-mediated breakage and reunion is the drug–DNA complex.

The results of computer simulation [8], based on formation constants of HNaI complexing with a range of metal ions, suggested that HNaI is mainly non-complexed in plasma, and so acts intracellularly. A mixed ligand complex between the drug, metal ion, and DNA (with the metal ion acting as a bridge) was proposed as a transition state. A study of the partition coefficients of nalidixic acid and of the complexes with Cu(II), Zn(II) and Mg(II) led to the conclusion that lipid solubility is not a major factor in the efficacy of either the parent drug or the 1:1 drug–metal complexes [9].

We report here the results of attempts to isolate some solid complexes of nalidixic acid with the aims of studying how HNaI binds to various metal ions and of obtaining materials for biological testing.

### Results and Discussion

Nalidixic acid is a monoprotic acid ( $pK_a = 5.94$ ) which is absorbed gastrointestinally [10], so its use in the anionic form increases its solubility and absorp-

\*Author to whom correspondence should be addressed.

TABLE I. Analytical Data for some Complexes of Nalidixic Acid (=HL)

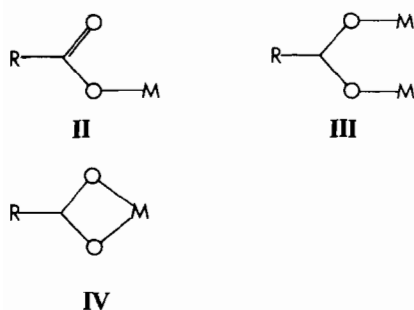
Complex	M:L <sup>a</sup>	Colour	Analysis (%)					
			Found			Calculated		
			C	H	N	C	H	N
Mg(L) <sub>2</sub> ·H <sub>2</sub> O	1:2	white	56.64	4.67	10.87	57.11	4.79	11.10
Ca(L)(OH)	1:2	white	49.44	4.07	9.49	50.00	4.20	9.72
Cr(L) <sub>2</sub> (OH)	1:3	green	53.69	4.49	9.97	54.24	4.36	10.54
Mn(L) <sub>2</sub>	1:2	yellow	55.55	4.25	10.82	55.72	4.29	10.83
Fe(L) <sub>3</sub> ·2H <sub>2</sub> O	1:3	yellow	55.17	4.26	10.75	55.05	4.75	10.70
Fe(L) <sub>2</sub> (OH)	1:3	red-brown	53.84	4.62	10.09	53.86	4.33	10.47
Ni(L)(NO <sub>3</sub> )·H <sub>2</sub> O	1:2	green	40.01	3.51	11.17	38.96	3.54	11.36
Ni(L) <sub>2</sub> ·3H <sub>2</sub> O	1:2	blue-green	50.40	4.63	9.78	50.11	4.91	9.74
Cu(L) <sub>2</sub> ·H <sub>2</sub> O	1:1	blue	52.84	4.11	10.14	52.98	4.45	10.30
Zn(L) <sub>2</sub> ·H <sub>2</sub> O	1:1	white	52.14	4.33	10.09	52.81	4.43	10.26
Pd(L)·H <sub>2</sub> O	1:2	brown	48.07	3.77	9.30	49.12	4.12	9.55
Cd(L) <sub>2</sub>	1:2	white	50.05	3.91	9.65	50.14	3.86	9.75
Cd(L)Br	1:1	white	33.83	2.57	6.44	34.03	2.62	6.61
Hg(L)Cl	1:2	white	31.56	2.65	6.68	30.84	2.37	6.00

<sup>a</sup>Ratio used in synthesis.

tion. For our study of the interaction of this antibiotic with metal ions we therefore chose to use the sodium salt as the starting ligand. The metal complexes we have prepared are listed in Table I. Unfortunately none of them was isolated in the form of crystals suitable for structure determination by X-ray diffraction methods, so information about their structures was obtained by spectral studies.

In the IR spectrum of HNaI the  $\nu(\text{C}=\text{O})$  stretching mode of the carboxylic acid group is observed as a band at  $1720\text{ cm}^{-1}$ . This disappears on deprotonation and in the sodium salt there are two new bands at  $1590$  and  $1392\text{ cm}^{-1}$ , the carboxylate antisymmetric ( $\nu_a$ ) and symmetric ( $\nu_s$ ) vibrations respectively.

The carboxylate ion usually coordinates to metal ions in one of three main ways (II–IV). Such differences are reflected in the relative positions of the



antisymmetric and symmetric stretching vibrations. The separations between the  $\nu_a$  and  $\nu_s$ ,  $\Delta\nu$ , in unidentate complexes (structure II) are expected to be

much larger than in the free ion. In a bidentate (chelate) complex (structure IV)  $\Delta\nu$  will be significantly smaller than in the free ion whereas in the bridging complex (structure III) the  $\Delta\nu$  value is closer to that of the free ion [11].

TABLE II.  $\nu(\text{OCO})$  Frequencies ( $\text{cm}^{-1}$ ) for the Sodium Salt of Nalidixic Acid (NaI) and some of its Complexes

Compound	$\nu(\text{OCO})$			Coordination mode
	$\nu_a$	$\nu_s$	$\Delta\nu$	
NaNaI	1590	1392	198	
Ca(NaI)OH	1583	1395	188	bridging
Mg(NaI) <sub>2</sub> ·H <sub>2</sub> O	1572	1402	170	bridging
Mn(NaI) <sub>2</sub>	1575	1388	187	bridging
Ni(NaI) <sub>2</sub> ·3H <sub>2</sub> O	1574	1398	176	bridging
Ni(NaI)NO <sub>3</sub> ·H <sub>2</sub> O	1570	1400	170	bridging
Zn(NaI) <sub>2</sub> ·H <sub>2</sub> O	1570	1390	180	bridging
Cd(NaI) <sub>2</sub>	1572	1385	187	bridging
Cr(NaI) <sub>2</sub> OH	1620	1495	125	chelate
Fe(NaI) <sub>3</sub> ·2H <sub>2</sub> O	1620	1483	137	chelate
Cu(NaI) <sub>2</sub> H <sub>2</sub> O	1610	1485	125	chelate
Pd(NaI) <sub>2</sub> H <sub>2</sub> O	1590	1485	105	chelate

The compounds obtained in the present work may be divided into two groups on the basis of their IR spectra (Table II): (a) those having bridging carboxylates and (b) those in which the carboxylate group chelates. In no case was evidence found for the keto group on C(4) coordinating to a metal ion as the band at  $1625\text{ cm}^{-1}$  due to the  $\nu(\text{C}=\text{O})$  stretch of this group in NaNaI remained invariant in the metal

complexes nor were any changes observed in the ketone deformation region (1200–1350  $\text{cm}^{-1}$ ).

Assignment of the type of carboxylate group coordination was based on both the position of the  $\nu_s$  band and the values of  $\Delta\nu$  [12–14]. In the compounds in group (a), in which the carboxylate group functions as a bridging ligand,  $\nu_s(\text{OCO})$  was found at 1385–1402  $\text{cm}^{-1}$  and  $\Delta\nu$  was 170–190  $\text{cm}^{-1}$ . Compounds in group (b), in which the carboxylate group acts as a chelate, have  $\Delta_s(\text{OCO})$  1483–1495  $\text{cm}^{-1}$  and  $\Delta\nu$  105–137  $\text{cm}^{-1}$ .

#### Type (a) Complexes

This class, in which the carboxylate unit functions as a bridging group, comprises the compounds  $\text{M}(\text{Nal})_2 \cdot \text{H}_2\text{O}$  ( $\text{M} = \text{Mg}$ , and  $\text{Zn}$ ),  $\text{M}(\text{Nal})_2$  ( $\text{M} = \text{Mn}$ ,  $\text{Cd}$ ),  $\text{Ni}(\text{Nal})_2 \cdot 3\text{H}_2\text{O}$ ,  $\text{Ni}(\text{Nal})(\text{NO}_3) \cdot \text{H}_2\text{O}$  and  $\text{Ca}(\text{Nal})(\text{OH})$ . All the compounds were apparently air-stable. They were insoluble in water and common organic solvents, except for  $\text{Ni}(\text{Nal})(\text{NO}_3)\text{H}_2\text{O}$  which dissolved in ethanol.

The reflectance spectra of the solid nickel complexes were typical of the presence of an octahedral ligand field of oxygen donor ligands (e.g. for  $\text{Ni}(\text{Nal})_2 \cdot 3\text{H}_2\text{O}$ :  $\nu_1$  at 8760  $\text{cm}^{-1}$ ,  $\nu_2 + {}^3\text{A}_{2g} \rightarrow {}^1\text{E}_g$  at 14050 and 14780  $\text{cm}^{-1}$ ).

The X-band EPR spectrum of a polycrystalline sample of  $\text{Mn}(\text{Nal})_2$  consisted of a single band at  $g = 2$  (15 mT width peak to peak). This is consistent with either a highly symmetrical monomeric ligand field or a polymeric structure with consequent dipolar interactions between the adjacent metal centres, resulting in the simple spectrum observed [15]. As the presence of carboxylate bridges may be inferred from the IR spectrum the latter, polymeric, structure seems more probable.

Cadmium forms a complex of the same stoichiometry,  $\text{Cd}(\text{Nal})_2$ , and the two compounds  $\text{M}(\text{Nal})_2$  ( $\text{M} = \text{Mn}$  and  $\text{Cd}$ ) have very similar  $\nu_a$  and  $\nu_s(\text{OCO})$

values. The EPR spectrum of a sample of  $\text{Cd}(\text{Nal})_2$  doped with manganese (nominal 1 mol %) (Fig. 1) consisted of a band at  $g = 2$  split into six hyperfine components ( $\text{Mn } I = 5/2$ ). This result shows that the symmetry of the immediate environment of the  $\text{Mn}(\text{II})$  ions in the  $\text{Cd}(\text{Nal})_2$  lattice is quite high, and an essentially tetrahedral geometry of oxygen atoms from bridging carboxylate groups is proposed. The slightly irregular form of the spectrum suggests the presence of a very small degree of zero-field splitting, reflecting a small departure from rigorously tetrahedral geometry. The spectrum of  $\text{Mn}(\text{Nal})_2$ , is consistent with a similar structure, the absence of hyperfine structure being due to the dipolar interaction between adjacent  $\text{Mn}(\text{II})$  ions in the polymer.

#### Type (b) Complexes

This class, for which the IR results suggest the presence of chelating carboxylate groups, comprises the compounds  $\text{Cu}(\text{Nal})_2 \cdot \text{H}_2\text{O}$ ,  $\text{Pd}(\text{Nal})_2 \cdot \text{H}_2\text{O}$ ,  $\text{Fe}(\text{Nal})_3 \cdot \text{H}_2\text{O}$ , and  $\text{Cr}(\text{Nal})_2\text{OH}$ .

The electronic spectrum of  $\text{Cu}(\text{Nal})_2 \cdot \text{H}_2\text{O}$  had a strong band at 15380  $\text{cm}^{-1}$  with a shoulder at 16700  $\text{cm}^{-1}$ , and its X-band EPR spectrum was of the axial type ( $g_{xy} 2.067$ ,  $g_z 2.173$ ), indicative of a tetragonal environment.

The reaction of chromium(III) chloride with the sodium salt of nalidixic acid in water gave a green compound, which after drying at 110  $^\circ\text{C}$ , had the formula  $\text{Cr}(\text{Nal})_2(\text{OH})$ . The EPR spectrum of the solid compound showed a broad (44.5 mT peak-to-peak breadth) band centred at  $g = 1.97$ , indicative of the presence of strong dipolar coupling between the paramagnetic ions, as commonly found for polymeric complexes.

It is well known that  $\text{Cr}(\text{III})$  in aqueous solution exhibits several hydrolysis equilibria [16]. Under the reaction conditions employed in the synthesis ( $[\text{Cr}] = 0.014 \text{ M}$ ,  $\text{pH} = 3.4$ ) hydroxo-bridged species would be present and would favour the formation of a dinuclear structure for  $\text{Cr}(\text{Nal})_2(\text{OH})$  as in V.

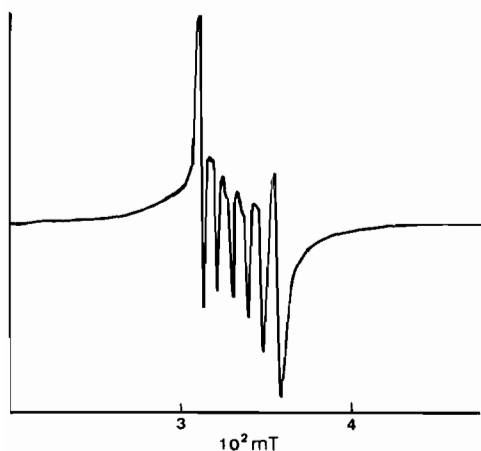
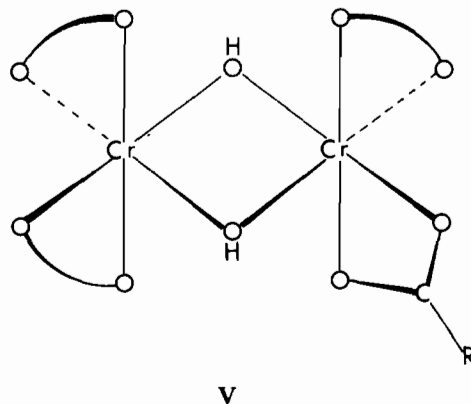


Fig. 1. X-Band EPR spectrum of  $\text{Cd}(\text{Mn})(\text{Nal})_2$ .



Two products were obtained from the reaction of iron(III) chloride and the sodium salt of nalidixic acid in water: yellow  $\text{Fe}(\text{Nal})_3 \cdot 2\text{H}_2\text{O}$  and red-brown  $\text{Fe}(\text{Nal})_2(\text{OH})$ . Under the reaction conditions employed ( $[\text{Fe}] = 0.025 \text{ m}$ ,  $\text{pH} = 2.1$ ) both mononuclear  $\text{Fe}(\text{H}_2\text{O})_6^{3+}$  and dinuclear hydrolysed complexes are the principal species [16], so the formation of more than one nalidixic acid complex is not unexpected.

The major product was  $\text{Fe}(\text{Nal})_3 \cdot 2\text{H}_2\text{O}$  and this had an X-band EPR spectrum consisting of a set of at least eight bands in the 0–0.6 T region (Fig. 2). Such spectra are expected [15] for high spin iron(III) species when the principal zero-field splitting parameter,  $D$ , is finite but small ( $<ca. 0.1 \text{ cm}^{-1}$ ), corresponding to only small deviations of the ligand field environment from regularly octahedral geometry.

This conclusion is supported by the Mössbauer spectrum of the complex (Fig. 3) which consisted of a single broad peak (unresolved doublet). The form of the spectrum and the chemical isomer shift value,  $\delta$ , (Table III) are very similar to those of the trioxalatoferate(III) anion,  $\text{Fe}(\text{C}_2\text{O}_4)_3^{3-}$  [17].

The Mössbauer (Fig. 4) and EPR (Fig. 5) spectra of the minor product of the reaction, red-brown  $\text{Fe}(\text{Nal})_2(\text{OH})$ , differ appreciably from those of  $\text{Fe}(\text{Nal})_3 \cdot 2\text{H}_2\text{O}$ . The quadrupole splitting observed

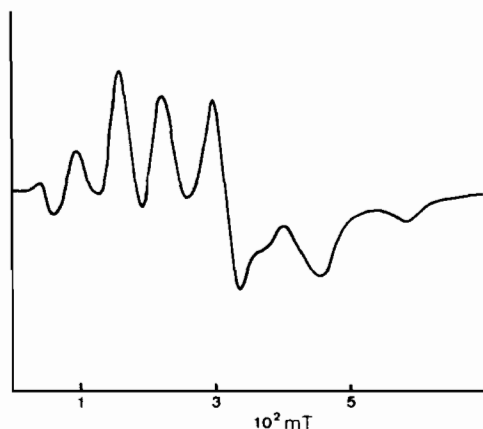


Fig. 2. X-band EPR spectrum of  $\text{Fe}(\text{Nal})_3 \cdot 2\text{H}_2\text{O}$ .

in the Mössbauer spectrum and the form of the EPR spectrum, notably the presence of the major band in the  $g \approx 4$  region, both indicate a more distorted environment about the iron atom than in  $\text{Fe}(\text{Nal})_3 \cdot 2\text{H}_2\text{O}$ . However, these results alone do not permit us reliably to assign a structure to  $\text{Fe}(\text{Nal})_2(\text{OH})$ .

In conclusion, it appears that, under the experimental conditions we have employed, a range of solid HNaI metal complexes may be isolated. In these the

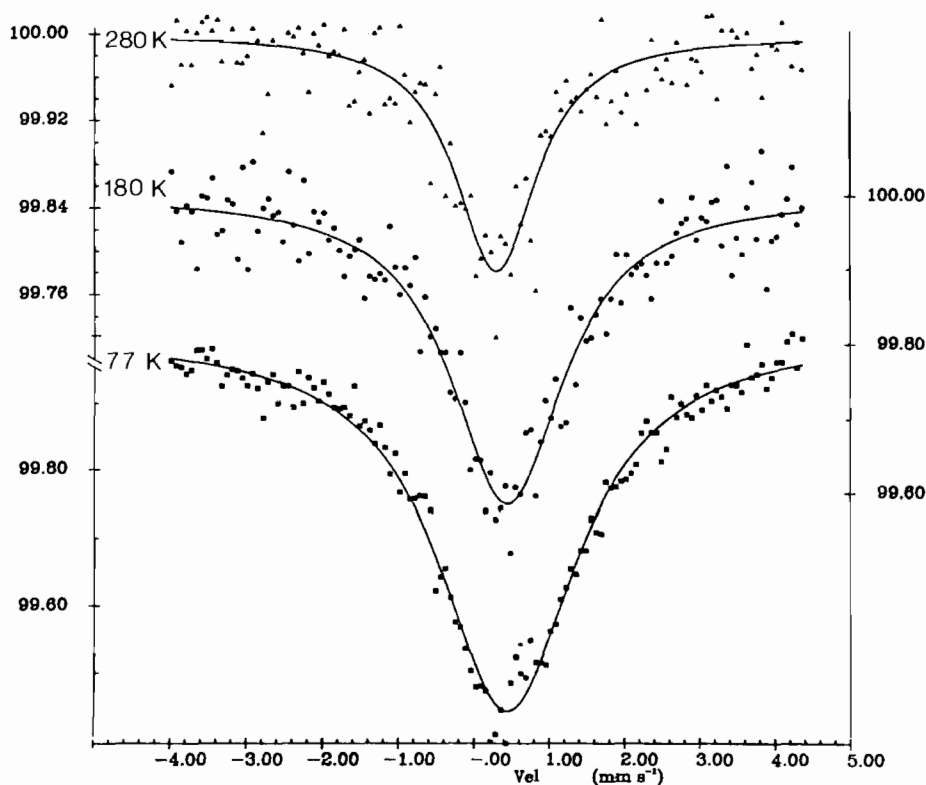


Fig. 3. Mössbauer spectrum of  $\text{Fe}(\text{Nal})_3 \cdot 2\text{H}_2\text{O}$ .

TABLE III. Mössbauer Parameters ( $\text{mm s}^{-1}$ ) of the Iron-(III) Complexes with Nalidixic Acid

Compound	$\delta^a$	$\Delta^b$	$\Gamma^c$	$T$ (K)
$\text{Fe}(\text{Nal})_3 \cdot 2\text{H}_2\text{O}$	0.44(2)		1.13(6)	80
	0.45(3)		0.91(9)	180
	0.29(4)		0.66(9)	280
$\text{Fe}(\text{Nal})_2\text{OH}$	0.466(6)	0.80(1)	0.28(1)	80
	0.427(4)	0.787(7)	0.263(7)	180
	0.361(4)	0.767(7)	0.259(6)	280

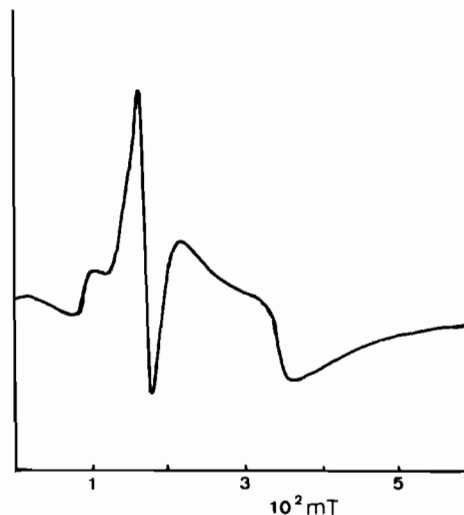
<sup>a</sup>Chemical isomer shift ( $^{57}\text{Fe}$  as reference). <sup>b</sup>Quadrupole splitting. <sup>c</sup>Signal width at half maximum.

nalidixic acid anion binds through the carboxylate group either as a chelate or as a bridge to give a polymeric structure. Studies of the biological activity of some of these compounds are currently in progress.

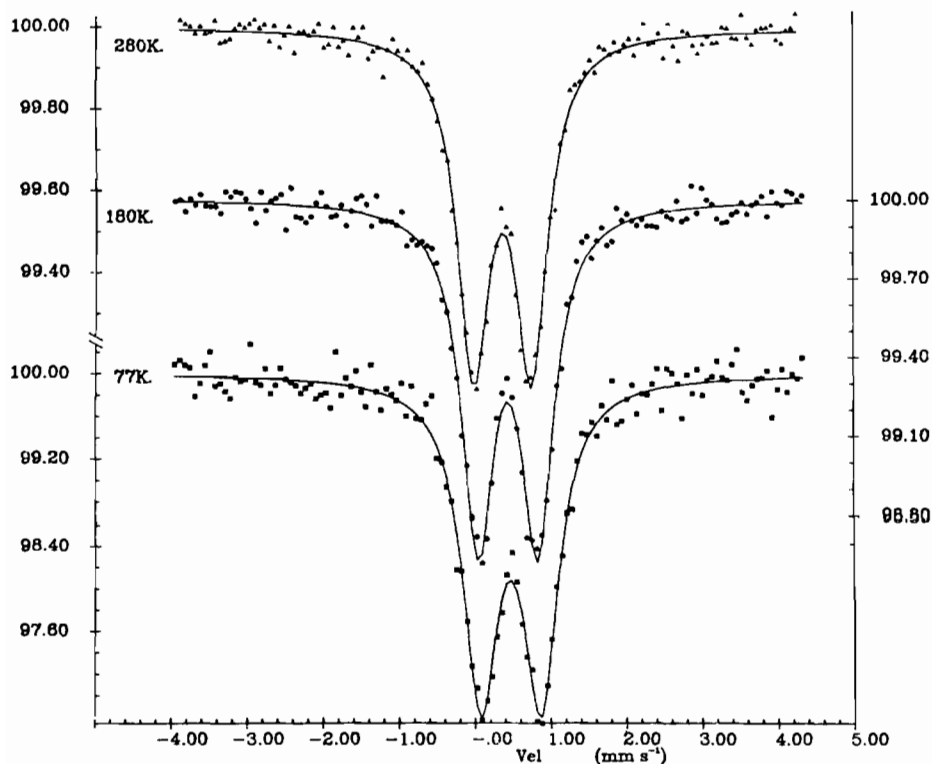
## Experimental

### Preparation

The sodium salt of nalidixic acid (NaNal) was prepared by mixing equimolar amounts of  $\text{NaHCO}_3$  and HNaI in aqueous solution. The reaction mixture was heated until clear and then used immediately.

Fig. 5. X-band EPR spectrum of  $\text{Fe}(\text{Nal})_2\text{OH}$ .

Except where stated otherwise, the complexes were prepared by the following general method. An aqueous solution of NaNal was added to one of the metal chloride in the stoichiometry ratios given in Table I. The complex usually precipitated immediately or within a short time after mixing the reactants. The products were washed with water and dried at  $100^\circ\text{C}$ .

Fig. 4. Mössbauer spectrum of  $\text{Fe}(\text{Nal})_2\text{OH}$ .

*Mn(Nal)<sub>2</sub>*

The reaction was carried out in hot water.

*[Fe(Nal)<sub>3</sub>]·2H<sub>2</sub>O and [Fe(Nal)<sub>2</sub>(OH)]*

Both products were obtained from the same reaction. The reaction mixture was concentrated to small volume and the precipitate obtained was recrystallized from CHCl<sub>3</sub>. The yellow solid which separated from the CHCl<sub>3</sub> solution was Fe(Nal)<sub>3</sub>·2H<sub>2</sub>O. The mother liquid subsequently precipitated red-brown Fe(Nal)<sub>2</sub>(OH).

*Ni(Nal)(NO<sub>3</sub>)·H<sub>2</sub>O*

The salt employed was Ni(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O and the reaction was carried out in ethanol.

*Ni(Nal)<sub>2</sub>·3H<sub>2</sub>O*

The salt employed was Ni(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O and the reaction was carried out in water.

*Pd(Nal)<sub>2</sub>·H<sub>2</sub>O*

The salt used was K<sub>2</sub>PdCl<sub>4</sub>.

*Cd(Nal)<sub>2</sub>*

This was prepared from Cd(NO<sub>3</sub>)<sub>2</sub> and NaNal by refluxing the reaction mixture with stirring for 6 h in ethanol.

*[Cd(Nal)Br<sub>2</sub>]*

The starting metal salt employed was CdBr<sub>2</sub>·4H<sub>2</sub>O.

*Cd(Mn)(Nal)<sub>2</sub>*

NaNal was added to an ethanolic solution of Cd(NO<sub>3</sub>)<sub>2</sub>·4H<sub>2</sub>O containing 1 mol% of MnCl<sub>2</sub>·4H<sub>2</sub>O, and the mixture was refluxed with stirring for 6 h.

Microanalyses (Table I) were by the Micro-analytical Laboratory, Imperial College.

*Physical Measurements*

Infrared spectra were obtained using Perkin-Elmer 681-B and 599-B spectrometers. Electronic spectra were measured by the diffuse reflectance technique on a Cary 14 instrument. X-Band EPR spectra were measured on polycrystalline samples using a Varian E-12 spectrometer. The Mössbauer spectra were

obtained on a standard spectrometer system by courtesy of Dr. L. V. C. Rees of the Physical Chemistry Section, Imperial College.

**Acknowledgements**

We thank the PSPA program of UNAM and CONACYT for the support of G.M.D. in his M.Sc. studies and research work, and the SERC for the EPR spectrometer. We are indebted to Dr. I. E. G. Morrison for the Mössbauer spectra. We thank the British Council for travel funds in support of the collaborative programme.

**References**

- 1 A. Albert 'Selective Toxicity. The Physico-Chemical Basis of Therapy', 6th edn., Chapman and Hall, London, 1979.
- 2 M. N. Hughes, 'The Inorganic Chemistry of Biological Processes', 2nd edn., Wiley, New York, 1981.
- 3 P. G. Sammes, in P. G. Sammes (ed.), 'Topics in Antibiotic Chemistry', Vol. 3, Halsted Press, New York, 1980.
- 4 N. R. Cozarelli, *Science*, 207, 953 (1980).
- 5 M. Gellert, *Ann. Rev. Biochem.*, 50, 879 (1981).
- 6 K. Timmers and R. Sternglanz, *Bioinorg. Chem.*, 9, 145 (1978).
- 7 L. L. Shen and A. G. Pernet, *Proc. Natl. Acad. Sci. U.S.A.*, 82, 307 (1985).
- 8 A. Cole, J. Goodfield and D. R. Williams, *Inorg. Chim. Acta*, 92, 91 (1984).
- 9 A. J. G. Bailey, A. Cole, J. Goodfield, P. M. May, M. E. Dreyfuss, J. M. Midgley and D. R. Williams, *Int. J. Pharm.*, 22, 283 (1984).
- 10 W. B. Pratt, 'Chemiotherapy of Infection', Oxford Univ. Press, Oxford, 1977.
- 11 K. Nakamoto, 'Infrared and Raman Spectra of Inorganic and Coordination Compounds', 3rd edn., Wiley, New York, 1977.
- 12 L. P. Battaglia, A. Bonamartini Corradi, G. Marcotrigiano, L. Menabue and G. C. Pellacani, *J. Am. Chem. Soc.*, 102, 2663 (1980).
- 13 L. P. Battaglia, A. Bonamartini Corradi, G. Marcotrigiano, L. Menabue and G. C. Pellacani, *Inorg. Chem.*, 20, 1075 (1981).
- 14 L. Antolini, L. Menabue, G. C. Pellacani, M. Saladini, L. P. Battaglia, A. Bonamartini Corradi and G. Marcotrigiano, *J. Chem. Soc., Dalton Trans.*, 2325 (1984).
- 15 R. D. Dowsing, J. F. Gibson, D. M. L. Goodgame, M. Goodgame and P. J. Hayward, *Nature (London)*, 219, 1037 (1968).
- 16 C. F. Baes and R. E. Mesmer, 'The Hydrolysis of Cations', Wiley, New York, 1976.
- 17 K. G. Dharmawardena and G. M. Bancroft, *J. Chem. Soc. A*, 2655 (1968).